






Cytotoxic Constituents from the Rhizomes of *Monstera deliciosa*

Marwa Elsbaey ¹, Kadria F. M. Ahmad ¹, Fathy A. Behery ^{1,2*},
Mohamed MA. Amer ¹ and Mohamed-Farid I. Lahloub ¹

¹ Department of Pharmacognosy, Faculty of Pharmacy, Mansoura University, Mansoura 35516, Egypt

² Department of Pharmacy, College of Pharmacy, Riyadh Elm University, Riyadh 11681, Saudi Arabia

(Received March 18, 2022; Revised August 07, 2022; Accepted August 09, 2022)

Abstract: A phytochemical study of the rhizomes of *Monstera deliciosa* has led to isolation of fourteen compounds (1-14) for the first time from the plant species. The structure elucidation was carried out using 1D and 2D NMR data. The cytotoxic activities of the compounds against the liver (HepG-2), larynx (Hep-2), colon (HCT-116), and breast (MCF-7) cancer cell lines were evaluated using MTT assay. Syringaresinol (6) showed remarkable IC₅₀ values against the four tested cell lines. In addition, 9, 12, 13-trihydroxy-10-octadecenoic acid (14) was highly cytotoxic against Hep-2 and HCT-116 cell lines.

Keywords: *Monstera*; Araceae; cytotoxic; lignan. © 2022 ACG Publications. All rights reserved.

1. Plant Source

Monstera deliciosa Liebm. whole plant was collected from Mansoura University, Mansoura, Egypt in July 2011, and the rhizomes of the species were separated from the rest of the plant. The plant was identified by Dr. Mahmoud Makram Kassem, Department of Vegetables and Ornamentals, Faculty of Agriculture, Mansoura University. A voucher specimen was deposited at the Department of botany and microbiology, Faculty of Science, Herbarium, Damietta University (DAM00022).

2. Previous Studies

In our previous *in vivo* study, the rhizomes of *M. deliciosa* exhibited significant antihyperglycemic effects in STZ-induced rats [1]. *In vitro* activity on the ethanol extract of the fruit of the species was shown to increase the insulin secretion [2]. Cytotoxic [3], anti-inflammatory, and wound healing activities of the extract of the leaves of the species were also reported in former studies. However, the extract of the leaves did not exhibit any antibacterial activity [4]. In addition to those activities, the *in vivo* anticancer and antioxidant activities of the methanol extract of the species were reported [5]. In contrast to the leaves of the species, the extract of stems was reported as an

* Corresponding author: E- Mail: fathybehery@mans.edu.eg (F. Behery), Phone +966-5069-89721.

antibacterial agent together with antioxidant activity [2]. The simple phenolics, flavonoids, and tannins contents of the species were reported from the stem of the plant [2]. About 400 volatile constituents of the fruit of the species were reported by simultaneous steam distillation-solvent extraction and GC-MS [7].

3. Present Study

The different fractions of the methanol extract the rhizomes of *M. deliciosa* were subjected to chromatographic separation and purification leading to the isolation of 14 compounds for the first time from the plant. The present study deals with the isolation and identification of those compounds together with their *in vitro* cytotoxic activities against the liver (HepG-2), larynx (Hep-2), colon (HCT-116) and breast (MCF-7) cancer cell lines. The dried rhizomes (3 kg) of *M. deliciosa* were powdered and extracted by maceration with MeOH (6 x 5 L) at room temperature (20-25 °C). The filtered and dried extract (200 g) was partitioned with H₂O/petroleum ether and H₂O/CH₂Cl₂ to afford the petroleum ether (45.5 g) and methylene chloride (14.7g) fractions, respectively. The resulting fractions were subjected to normal (Silica Gel G 60-230, Merck, Germany) and reversed-phase chromatography (RP-C₁₈, Merck, Germany) as well as crystallization to afford compounds **2**, **4**, **5**, **9**, **10**, **11**, **12**, and **13** from the petroleum ether fraction and compounds **1**, **2**, **6**, **7**, **8**, and **14** from the methylene chloride fraction. Detailed purification steps are depicted in Figures S1-S4.

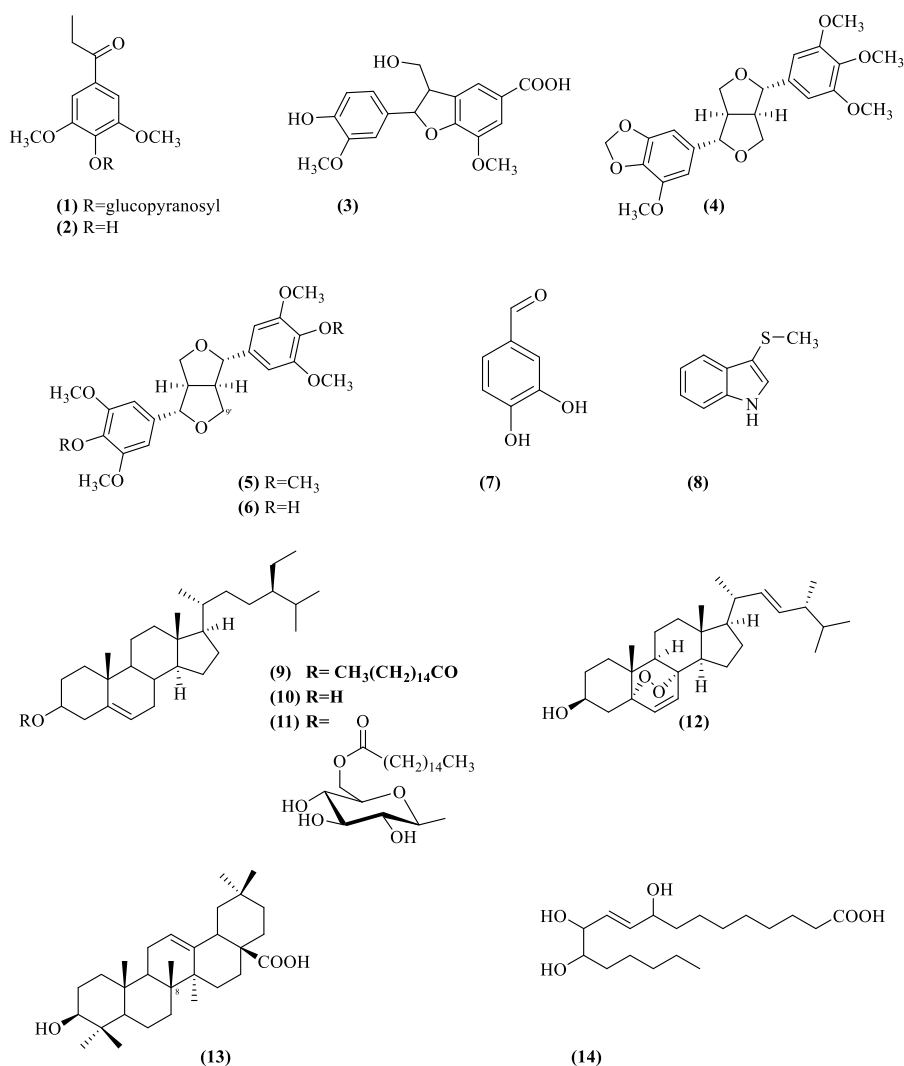


Figure 1. Structures of compounds (1-14) isolated from the rhizomes of *M. deliciosa*

Cytotoxic Constituents from Rhizomes of *Monstera deliciosa*

Cytotoxic activity- MTT Assay: The assay was carried out according to Mauceri [8]. The cytotoxic activity was investigated against human cancer cell lines from the liver (HepG-2), larynx (Hep-2), colon (HCT-116), and breast (MCF-7) originated from ATCC (Manassas, VA, USA) and were obtained from VACSERA, Cairo, Egypt. The isolated compounds were dissolved in DMSO and diluted with PBS to concentrations of 400, 200, 100, 50, and 25 µg/mL. In all experiments, control cells were dissolved with DMSO alone. A BioTeck® microplate reader (Winooski, VT, USA) was used to determine the optical densities. Statistical analysis of the data was performed using Microsoft Excel software version 2010.

Compounds **1-14** were isolated using a series of normal and reversed-phase chromatography as well as crystallization. The compounds were identified based on their NMR spectroscopic data (Figure 1). Their structures were confirmed by comparing their spectral data with those reported in the literature. They were identified as follows; propiosyringone- β -D-glucopyranoside (**1**) [9], propiosyringone (**2**) [10], ceplignan (**3**) [11], sesartemin (**4**) [12], yangambin (**5**) [12], syringaresinol (**6**) [13], protocatechuic aldehyde (**7**) [14], 3-methyl thio-indole (**8**) [15], β -sitosteryl palmitate (**9**) [16], β -sitosterol (**10**) [17], 7-oxo- β -sitosterol-3-O- β -D-glucopyranoside)-6'-palmitate (**11**) [18], 5 α , 8 α -epi-dioxyergosta-6, 22-dien-3 β -ol (**12**) [19], oleanolic acid (**13**) [19], 9, 12, 13-trihydroxy-10-octadecenoic acid (**14**) [20]. This is the first report of compounds **1-14** from *M. deliciosa* (Figures S5-S39, Tables S1-S8).

The cytotoxic activities of compounds **1-14** were evaluated using an MTT assay against human cancer cell lines from the liver (HepG-2), larynx (Hep-2), colon (HCT-116), and breast (MCF-7) (Table 1 and see S1, in supporting information) [8]. For HepG-2 cells, compound **6** showed the highest cytotoxicity with an IC₅₀ value (19.01 ± 0.16 µM) that is about a third of that of 5-Fu (IC₅₀ = 62.57 ± 1.85 µM). Meanwhile, compounds **9** (IC₅₀ = 31.53 ± 0.81 µM) and **14** (IC₅₀ = 30.50 ± 0.43 µM) showed remarkable activity as about half the IC₅₀ value of 5-FU. It is worth mentioning that β -sitosterol is reported to exert antihepatocellular cancer activity [21]. It was noticed that esterification with palmitate greatly enhanced the activity since compound **10** showed IC₅₀ value of 55.55 ± 0.79 µM, which is still higher than 5-FU. All compounds showed comparable or higher cytotoxic activity on Hep-2 cells as compared to 5-FU (IC₅₀ = 40.12 ± 0.62 µM) except for compound **8** (IC₅₀ = 71.85 ± 1.32 µM) which was about half the potency of 5-FU.

Table 1. Cytotoxicity data of compounds **1-14** and 5-Fluorouracil (**5-FU**) against different cell lines using MTT assay as IC₅₀ values

Compounds	<i>In vitro</i> Cytotoxicity IC ₅₀ (µM)			
	HepG2	Hep-2	HCT-116	MCF-7
1	65.86 ± 2.02	24.64 ± 0.53	30.14 ± 0.64	66.34 ± 1.18
2	53.83 ± 0.94	23.7 ± 0.62	31.65 ± 0.46	55.88 ± 0.96
4	44.86 ± 0.9	18.16 ± 0.59	21.67 ± 0.78	64.44 ± 0.02
5	71.35 ± 2.03	41.18 ± 0.4	28.53 ± 0.25	71.98 ± 0.35
6	19.01 ± 0.16	10.3 ± 0.47	14.34 ± 0.46	16.59 ± 0.17
7	45.03 ± 2.19	29.68 ± 0.11	44.81 ± 0.85	45.82 ± 0.98
8	128.6 ± 3.81	71.85 ± 1.32	60.89 ± 0.37	122.8 ± 1.72
9	31.53 ± 0.81	39.3 ± 1.03	12.43 ± 0.62	32.2 ± 0.71
10	55.55 ± 0.79	23.24 ± 1.14	25.68 ± 1.63	53.91 ± 1.02
12	53.93 ± 0.81	19.87 ± 0.19	24.98 ± 0.87	48.36 ± 0.01
13	60.12 ± 1.18	23.66 ± 0.88	23.49 ± 0.88	66.06 ± 0.11
14	30.5 ± 0.43	11.38 ± 0.67	16.18 ± 0.58	28.88 ± 0.21
5-FU	62.57 ± 1.85	40.12 ± 0.62	48.04 ± 0.39	38.05 ± 1.22

Data are expressed as mean ± SD of three independent experiments, each done in triplicate.

While compounds **6** and **14** were the highest cytotoxic with IC₅₀ values of 10.3 ± 0.47 and 11.38 ± 0.67 µM, respectively, about fourth that of 5-FU (IC₅₀ = 40.12 ± 0.62 µM) against Hep-2 cells, the

compounds **10**, **13**, **2** and its glucoside **1** showed comparable activity with IC₅₀ values of 10.3 ± 0.47, 23.66 ± 0.88, 23.7 ± 0.62 and 24.64 ± 0.53 µM, respectively, corresponding to about half of the IC₅₀ value of 5-FU. Cytotoxicity properties of oleanolic acid (**13**) against Hep-2 cells are previously reported in the literature [22].

For HCT-116, compounds **9**, **6**, and **14** showed significant cytotoxic activities with IC₅₀ values of 12.43 ± 0.62, 14.34 ± 0.46 and 16.18 ± 0.58 µM, respectively, as compared to 5-FU (IC₅₀ = 48.04 ± 0.39 µM).

While compound **6** showed the highest cytotoxicity against MCF-7 cell line ((IC₅₀ = 16.59 ± 0.17 µM) as compared to 5-FU (IC₅₀ = 38.05 ± 1.22 µM), compounds **9** and **14** also exhibited remarkable activity against the cell line as 32.2 ± 0.71 and 28.88 ± 0.21 µM, respectively.

Acknowledgments

Special thanks to Prof. Dr. Khaled A. El Sayed, School of Basic Pharmaceutical and Toxicological Sciences, College of Pharmacy, University of Louisiana at Monroe, Monroe, Louisiana 71201, United States, for the help in conducting many of the spectral analysis. The authors would like to Acknowledge Prof. Dr. Sami Rabei, Professor of Plant Taxonomy and Flora, Botany and Microbiology Department, Faculty of Science, Damietta, for his kind efforts in preparing the plant specimen for the herbarium and for providing the herbarium number of the plant sample.

Supporting Information

Supporting Information accompanies this paper on <http://www.acgpubs.org/journal/records-of-natural-products>

ORCID

Marwa Elsbaey: [0000-0002-5781-9434](https://orcid.org/0000-0002-5781-9434)

Kadria F. M. Ahmad: [0000-0003-4680-6133](https://orcid.org/0000-0003-4680-6133)

Fathy Behery: [0000-0001-5889-034x](https://orcid.org/0000-0001-5889-034x)

Mohamed MA Amer [0000-0002-5894-8641](https://orcid.org/0000-0002-5894-8641)

Mohamed-Farid Lahloub: [0000-0001-6769-3145](https://orcid.org/0000-0001-6769-3145)

References

- [1] M. Elsbaey, A. Awad, K. Fawzy, M. Amer and M. F. Lahloub (2014). Antidiabetic and antihyperlipidemic evaluation of *Monstera deliciosa* against streptozocin-induced diabetic rat, *J. Pharm. Phytother.* **2**, 1-4.
- [2] Z. Hussain, A. Waheed, R. A. Qureshi, D. K. Burdi, E. J. Verspohl, N. Khan and M. Hasan (2004). The effect of medicinal plants of Islamabad and Murree region of Pakistan on insulin secretion from INS-1 cells, *Phytotherapy Res.* **18**, 73–77.
- [3] D. N. Lira, M. A. Uddin, M. Uddin and A. S. S. Rouf (2014). Assessment of cytotoxic activities of *Phyllanthus amarus* and *Monstera deliciosa*, *J. App. Pharm. Science* **4**, 110–113.
- [4] M. Manirujjaman (2019). Identification of bioactive compounds in native plants and their effects on wound healing. PhD thesis, School of Clinical Sciences, Faculty of Health, Queensland University of Technology, Australia.
- [5] P. Prosanta, C. Mainak, K. Indrajit, H. Sagnik, D. Avratanu and H. P. Kanti (2015). Evaluation of anticancer activity of methanol extract of *Monstera deliciosa* in EAC induced Swiss albino mice, *Int. J. Toxicol. Pharmacol. Res.* **7**, 165-170
- [6] V. U. Rao, V. Viteesha, K. Suma and P. Nagababu (2015). Evaluation of phytochemical constituents, antibacterial and antioxidant activities of *Monstera deliciosa* Liebm. stem extracts, *World J. Pharm. Pharm. Sci.* **4**, 1422-1433.

Cytotoxic Constituents from Rhizomes of *Monstera deliciosa*

- [7] T. L. Peppard (1992). Volatile flavor constituents of *Monstera deliciosa*, *J. Agric. Food Chem.* **40**, 257–262.
- [8] H. J. Mauceri, N. N. Hanna, M. A. Beckett, D. H. Gorski, M. J. Staba, K. A. Stellato, K. Bigelow, R. Heimann, S. Gately and M. Dhanaba et al. (1998). Combined effects of angiostatin and ionizing radiation in antitumour therapy, *Nature* **394**, 287-291.
- [9] Y. F. Su, D. A. Guo, Y. J. Cui, J. S. Liu and J. H. Zheng (2001). A new phenolic glycoside and a new trans-clerodane diterpene from *Conyza blinii*, *J. Asian Nat. Prod. Res.* **3**, 229-233.
- [10] C. Zdero, F. Bohlmann, R. M. King and H. Robinson (1986). Diterpene glycosides and other constituents from *Argentinian baccharis* species, *Phytochemistry* **25**, 2841-2855.
- [11] M. Wen-Li, D. Hao-Fu and W. Da-Gang (2006). A new norneolignan from *Cephalomappa sinensis*, *Chem. J. Chinese U.* **27**, 1480-1481.
- [12] W. D. MacRae and G. N. Towers (1985). Non-alkaloidal constituents of *Virola elongata* bark, *Phytochemistry* **24**, 561-566.
- [13] X. Zhang, J. K. Xu, N. L. Wang, H. Kurihara and X. S. Yao (2008). Antioxidant phenanthrenes and lignans from *Dendrobium nobile*, *J. Chin. Pharm. Sci.* **17**, 314-318.
- [14] I. C. Lee, J. S. Bae, T. Kim, O. J. Kwon and T. H. Kim (2011). Polyphenolic constituents from the aerial parts of *Thymus quinquecostatus* var. japonica collected on Ulleung island, *J. Korean Soc. Appl. Biol. Chem.* **54**, 811-816.
- [15] M. H. Lee, Y. Y. Chen, J. W. Tsai, S. C. Wang, T. Watanabe and Y. C. Tsai (2011). Inhibitory effect of β -asarone, a component of *Acorus calamus* essential oil, on inhibition of adipogenesis in 3T3-L1 cells, *Food chem.* **126**, 1-7.
- [16] H. Sun, J. Zhang, Y. Ye, Y. Pan and Y. Shen (2003). Cytotoxic pentacyclic triterpenoids from the rhizome of *Astilbe chinensis*, *Helv. Chim. Acta.* **86**, 2414-2423.
- [17] D. D. Raga, C. L. Cheng, K. C. Lee, W. Z. Olaziman, V. J. De Guzman and C. C. Shen (2011). Bioactivities of triterpenes and a sterol from *Syzygium samarangense*, *Z. Naturforsch. C. Biosci.* **66**, 235-244.
- [18] R. M. Silverstein and G. C. Bassler (1962). Spectrometric identification of organic compounds, *J. Chem. Educ.* **39**, 546-553.
- [19] I. E. Díaz, E. G. Gonçalves and M. Yoshida. (2011). Constituyentes químicos del túbero de *Dracontium spruceanum* (Schott) G. Zhu ex *Dracontium lorentense* Krause (Araceae), *Bol. Soc. Quím. Peru.* **77**, 117-126.
- [20] J. Rösecke and W. A. König. Constituents of the fungi *Daedalea quercina* and *Daedaleopsis confragosa* var. Tricolor (2000), *Phytochemistry* **54**, 757-762.
- [21] T. K. Vo, Q. T. Ta, Q. T. Chu, T. T. Nguyen and V. G. Vo (2020). Anti-hepatocellular-cancer activity exerted by β -sitosterol and β -sitosterol-glucoside from *Indigofera zollingeriana* Miq, *Molecules* **25**, 3021-3034.
- [22] S. Kim, H. Lee, S. Lee, Y. Yoon and K. H. Choi (2015). Antimicrobial action of oleanolic acid on *Listeria monocytogenes*, *Enterococcus faecium*, and *Enterococcus faecalis*, *PLoS One* **10**, e0118800.

ACG
publications

© 2022 ACG Publications

© 2023. Notwithstanding the ProQuest Terms and Conditions, you may use this content in accordance with the associated terms available at <http://www.acgpubs.org/journal/records-of-natural-products/advisory-board>